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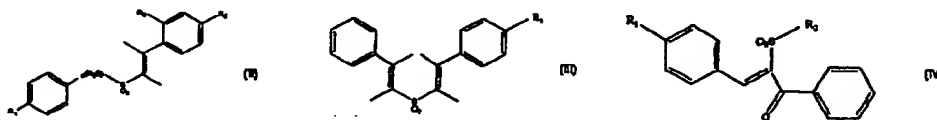
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## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(54) Title: **STYRYL SULFONE ANTICANCER AGENTS**

## (57) Abstract

Styryl sulfone compounds of the invention selectively inhibit proliferation of breast and prostate tumor cells, and induce apoptosis of such tumor cells, while sparing normal cells. The compounds, which are useful in the treatment of breast or prostate cancer, have formula (II) wherein *n* is zero or one; *R*<sub>1</sub> is selected from the group consisting of hydrogen, chlorine, fluorine and bromine; *R*<sub>2</sub> is selected from the group consisting of hydrogen, chlorine, fluorine, bromine, methyl and methoxy; and *R*<sub>3</sub> is selected from the group consisting of hydrogen, chlorine and fluorine; provided, *R*<sub>2</sub> may not be methyl or methoxy when *R*<sub>1</sub> and *R*<sub>3</sub> are both hydrogen and *n* is zero or one; and *R*<sub>1</sub>, *R*<sub>2</sub> and *R*<sub>3</sub> may not all be hydrogen when *n* is one, or formula (III) wherein *R*<sub>1</sub> is selected from the group consisting of hydrogen, chlorine, fluorine and bromine or formula (IV) wherein *R*<sub>1</sub> is selected from the group consisting of fluorine and bromine, and *R*<sub>2</sub> is selected from the group consisting of 2-chlorophenyl, 4-chlorophenyl, 4-fluorophenyl and 4-nitro.

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## **STYRYL SULFONE ANTICANCER AGENTS**

### **Field of the Invention**

The invention relates to compositions and methods for the treatment  
5 of cancer, in particular breast and prostate cancer.

### **Background of the Invention**

Extracellular signals received at transmembrane receptors are relayed into the cells by the signal transduction pathways (Pelech *et al.*, *Science* 257:1335 (1992)) which have been implicated in a wide array of physiological  
10 processes such as induction of cell proliferation, differentiation or apoptosis (Davis *et al.*, *J. Biol. Chem.* 268:14553 (1993)). The Mitogen Activated Protein Kinase (MAPK) cascade is a major signaling system by which cells transduce extracellular cues into intracellular responses (Nishida *et al.*, *Trends Biochem. Sci.* 18:128 (1993); Blumer *et al.*, *Trends Biochem. Sci.* 19:236 (1994)). Many steps of this  
15 cascade are conserved, and homologous for MAP kinases have been discovered in different species.

In mammalian cells, the Extracellular-Signal-Regulated Kinases (ERKs), ERK-1 and ERK-2 are the archetypal and best-studied members of the MAPK family, which all have the unique feature of being activated by  
20 phosphorylation on threonine and tyrosine residues by an upstream dual specificity kinase (Posada *et al.*, *Science* 255:212 (1992); Biggs III *et al.*, *Proc. Natl. Acad. Sci. USA* 89:6295 (1992); Garner *et al.*, *Genes Dev.* 6:1280 (1992)).

Recent studies have identified an additional subgroup of MAPKs, known as c-Jun NH2-terminal kinases 1 and 2 (JNK-1 and JNK-2), that have  
25 different substrate specificities and are regulated by different stimuli (Hibi *et al.*, *Genes Dev.* 7:2135 (1993)). JNKs are members of the class of stress-activated

protein kinases (SPKs). JNKs have been shown to be activated by treatment of cells with UV radiation, pro-inflammatory cytokines and environmental stress (Derijard *et al.*, *Cell* 1025 (1994)). The activated JNK binds to the amino terminus of the c-Jun protein and increases the protein's transcriptional activity by phosphorylating it at ser63 and ser73 (Adler *et al.*, *Proc. Natl. Acad. Sci. USA* 89:5341 (1992); Kwok *et al.*, *Nature* 370:223 (1994)).

Analysis of the deduced primary sequence of the JNKs indicates that they are distantly related to ERKs (Davis, *Trends Biochem. Sci.* 19:470 (1994)). Both ERKs and JNKs are phosphorylated on Tyr and Thr in response to external stimuli resulting in their activation (Davis, *Trends Biochem. Sci.* 19:470 (1994)). The phosphorylation (Thr and Tyr) sites, which play a critical role in their activation are conserved between ERKs and JNKs (Davis, *Trends Biochem. Sci.* 19:470 (1994)). However, these sites of phosphorylation are located within distinct dual phosphorylation motifs: Thr-Pro-Tyr (JNK) and Thr-Glu-Tyr (ERK). Phosphorylation of MAPKs and JNKs by an external signal often involves the activation of protein tyrosine kinases (PTKs) (Gille *et al.*, *Nature* 358:414 (1992)), which constitute a large family of proteins encompassing several growth factor receptors and other signal transducing molecules.

Protein tyrosine kinases are enzymes which catalyze a well defined chemical reaction: the phosphorylation of a tyrosine residue (Hunter *et al.*, *Annu Rev Biochem* 54:897 (1985)). Receptor tyrosine kinases in particular are attractive targets for drug design since blockers for the substrate domain of these kinases is likely to yield an effective and selective antiproliferative agent. The potential use of protein tyrosine kinase blockers as antiproliferative agents was recognized as early as 1981, when quercetin was suggested as a PTK blocker (Graziani *et al.*, *Eur. J. Biochem.* 135:583-589 (1983)).

The best understood MAPK pathway involves extracellular signal-regulated kinases which constitute the Ras/Raf/MEK/ERK kinase cascade (Boudewijn *et al.*, *Trends Biochem. Sci.* 20, 18 (1995)). Once this pathway is activated by different stimuli, MAPK phosphorylates a variety of proteins including several transcription factors which translocate into the nucleus and activate gene

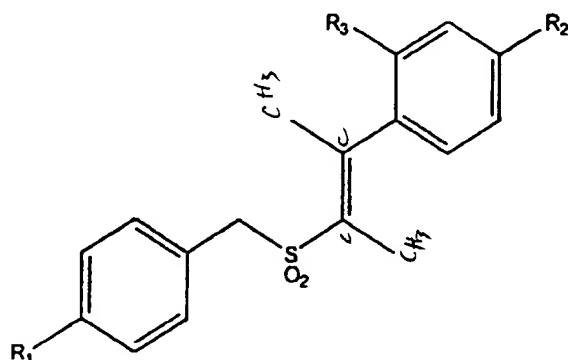
- 3 -

transcription. Negative regulation of this pathway could arrest the cascade of these events.

What are needed are new anticancer chemotherapeutic agents which target receptor tyrosine kinases and which arrest the Ras/Raf/MEK/ERK kinase cascade. Oncoproteins in general, and signal transducing proteins in particular, are likely to be more selective targets for chemotherapy because they represent a subclass of proteins whose activities are essential for cell proliferation, and because their activities are greatly amplified in proliferative diseases.

### Summary of the Invention

According to one embodiment of the invention, novel compounds are provided according to formula I:



wherein

$R_1$  and  $R_2$  are independently selected from the group consisting of chlorine, fluorine and bromine; and

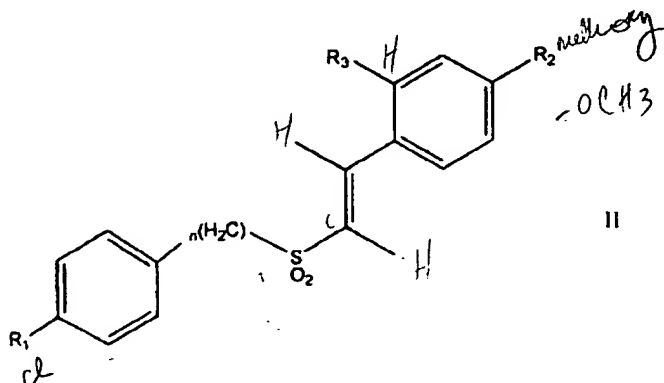
$R_3$  is selected from the group consisting of hydrogen and fluorine;

$R_1$  and  $R_2$  may not both be chlorine when  $R_3$  is hydrogen; and

$R_1$  may not be chlorine when  $R_2$  is fluorine and  $R_3$  is hydrogen in the same compound.

- 4 -

According to another embodiment of the invention, a pharmaceutical composition is provided comprising a pharmaceutically acceptable carrier and a compound of formula II



wherein

5

$n$  is zero or one;

$R_1$  is selected from the group consisting of hydrogen, chlorine, fluorine and bromine;

$R_2$  is selected from the group consisting of hydrogen, chlorine, fluorine, bromine, methyl and methoxy; and

10

$R_3$  is selected from the group consisting of hydrogen, chlorine and fluorine;

provided,

$R_2$  may not be methyl or methoxy when  $R_1$  and  $R_3$  are both hydrogen and  $n$  is zero or one; and

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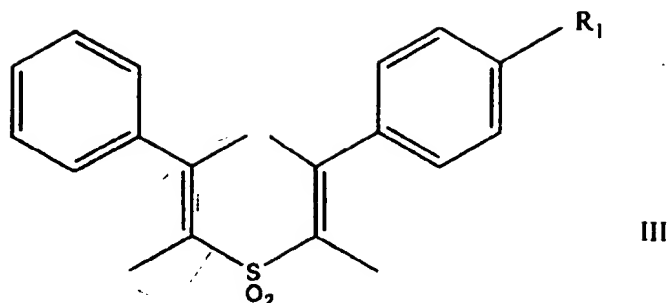
$R_1$ ,  $R_2$  and  $R_3$  may not all be hydrogen when  $n$  is one.

According to a preferred embodiment, the pharmaceutical composition comprises a pharmaceutically acceptable carrier and a compound of the formula II, wherein  $R_3$  is hydrogen, and  $R_1$  and  $R_2$  are independently selected from the group consisting of chlorine, fluorine and bromine.

20

According to another embodiment of the invention, a pharmaceutical composition is provided comprising a pharmaceutically acceptable carrier and a compound of the formula III

- 5 -

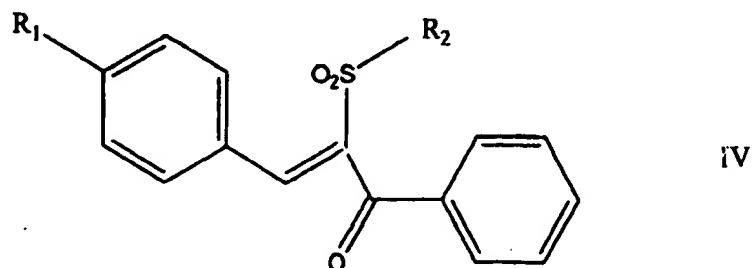


wherein

$R_1$  is selected from the group consisting of hydrogen, chlorine, fluorine and bromine.

According to another embodiment of the invention, a method of  
 5 treating an individual for breast or prostate cancer is provided, comprising  
 administering to said individual an effective amount of a compound according to  
 formula II or formula III, alone or in combination with a pharmaceutically acceptable  
 carrier. In another embodiment, a method of inhibiting growth of breast or prostate  
 tumor cells in an individual afflicted with breast or prostate cancer is provided,  
 10 comprising administering to said individual an effective amount of a compound  
 according to formula III, alone or in combination with a pharmaceutically acceptable  
 carrier. Furthermore, a method of inducing apoptosis of breast or prostate tumor cells  
 in an individual afflicted with breast or prostate cancer is provided comprising  
 administering to said individual an effective amount of a compound according to  
 15 formula III, alone or in combination with a pharmaceutically acceptable carrier.

The invention also relates to a pharmaceutical composition and  
 therapeutic methods as described above, wherein the compound is of the formula IV:





wherein

$R_1$  is selected from the group consisting of fluorine and bromine, and  $R_2$  is selected from the group consisting of 2-chlorophenyl, 4-chlorophenyl, 4-fluorophenyl and 4-nitro.

5

### Description of the Figures

Figs. 1A and 1B are bar graphs of the effect of compounds E-2,4-difluorostyryl-4-fluorobenzyl sulfone (FRI-2), E-4-fluorostyryl 4-bromobenzyl sulfone (FRI-6), E-4-bromostyryl 4-fluorobenzyl sulfone (FRI-7), E-4-fluorostyryl 4-chlorobenzyl sulfone (FRI-20) and E-4-chlorostyryl 4-chlorobenzyl sulfone (FRI-22) on NIH3T3, MCF7, BT-20 and LnCaP cells. Cells were treated with the compounds at 2.5  $\mu$ M (Fig. 1A) or 5.0  $\mu$ M (Fig. 1B) concentration and cell viability was determined after 48 hours by Trypan blue exclusion method. Each data point represents the average of three independent experiments. The standard deviation did not exceed 10%.

15

Fig. 2A is a bar graph of the concentration dependent inhibition of MCF7, BT20, LnCaP and NIH3T3 cells by treatment with FRI-20. The cells were treated with 0, 250 nM, 500 nM, 1  $\mu$ M, 2.5  $\mu$ M and 5.0  $\mu$ M FRI-20 for 48 hours. The percentage of living cells was determined by Trypan blue exclusion. The mean of three independent experiments is shown.

20

Fig. 2B is a bar graph of the viability of MCF7, BT20, LnCaP and NIH3T3 cells after treatment with FRI-20 at different time periods. All the cells were treated with FRI-20 at 2.5  $\mu$ M, and the number of viable cells was determined at 12, 24, 48, and 72 hours by Trypan blue exclusion. The mean of three independent experiments is shown.

25

Fig. 3A is a plot of the activity of compound FRI-20 on the normal cell lines NIH3T3, HeLa and HFL; the estrogen receptor-positive breast tumor cell lines MCF-7 and 361; the estrogen receptor-negative breast tumor cell lines SKBR-3, 435 and BT-20). Fig. 3B is similar to Fig. 3A except the treated cells comprise the androgen-dependent prostate cell line LnCaP, and the androgen independent prostate cell lines DU-145 and PC-3). All cells were treated with 2.5 and 5.0  $\mu$ M

30

concentration of FRI-20 and assayed for cell viability after 48 hours by Trypan blue exclusion. The mean of three experiments is shown. Variance did not exceed 10%.

Fig. 4 comprises a series of blots of the cell cycle analysis of FRI-20-treated or control-treated LnCaP cells. LnCaP cells were treated with 120 ml of DMSO (control cells) or 2.5  $\mu$ M FRI-20 in 10 ml of DMSO. Cells were harvested 6, 12, 24 and 48 hours following treatment and stained with propidium iodide and subjected to flow cytometry.

Fig. 5 is an SDS-PAGE autoradiograph of the effect of FRI-20 on ERK/MAPK activity. FRI-20-treated LnCaP, MCF-7 and NIH3T3 cells, along with DMSO-treated cells (control), were processed for ERK/MAPK immune complex kinase assay using myelin basic protein (MBP) as a substrate. The activity of ERK-2 toward MBP was then assayed in the presence of [ $\gamma^{32}$ P]ATP. The phosphorylated MBP was separated on 12% SDS-PAGE and visualized by autoradiography.

Fig. 6 is blot of the distribution of ERK-2 and JNK/SAPK proteins in NIH3T3, LnCaP and MCF-7 cells. Lysates of cultured cells containing 100 mg of proteins were loaded per lane. Following electrophoresis and transfer to a polyvinylidene membrane, proteins were blotted against ERK-2 and JNK-2 polyclonal antibodies and visualized by chemiluminescence.

Fig. 7 is an SDS-PAGE autoradiograph of the effect of FRI-20 on JNK/SAPK activity. JNK was immunoprecipitated from 100 mg of cultured cell lysates with JNK polyclonal antibody, and an immune complex kinase assay was carried out using GST-c-Jun (1-79) as a substrate. The phosphorylated proteins were separated by SDS-PAGE and visualized by autoradiography. The experiment was repeated three times with similar results.

## **Detailed Description of the Invention**

According to the present invention, certain styryl sulfone derivatives affect the MAPK signal transduction pathway, thereby affecting tumor cell growth and viability. The compounds inhibit the growth and proliferation of breast and prostate tumor cells in a dose-dependent manner, without affecting normal cell growth. This cell growth inhibition is associated with regulation of the ERK and JNK types of MAPK. The ability of the styryl sulfones to regulate these MAPKs and

- 8 -

induce cell growth arrest is dictated by the nature and position of the functional groups present in the compound.

Treatment of breast and prostate tumor cells with the styryl sulfone compounds of the invention leads to inhibition of cell proliferation and induction of apoptotic cell death. The effect is observed for estrogen receptor (ER) positive as well as estrogen receptor negative cells, although once breast cancer cell line tested, cell line 361, showed considerable resistance to styryl sulfones. Inhibition of cell proliferation and induction of apoptotic cell death is also observed for androgen-dependent as well as androgen-independent prostate tumor cells, although the former are considerably more sensitive to the styryl sulfones.

Tumor cells treated with the compounds of the invention accumulate in the G2/M phase of the cell cycle. As the cells exit the G2/M phase, they appear to undergo apoptosis. Treatment of normal cells with the styryl sulfones fails to produce a similar effect on cell cycle progression. Normal cells exhibit normal cell cycle progression in the presence and absence of styryl sulfone drug.

Both cells treated with the styryl sulfone compounds of the invention and untreated cells exhibit similar levels of intracellular ERK-2, but the biochemical activity of ERK-2, as judged by its ability to phosphorylate the substrate myelin basic protein (MBP), is considerably diminished in drug-treated cell compared to untreated cells. In prostate tumor cells, FR-20, a preferred compound of the invention, reduced the phosphorylation status of MBP by more than 80% compared to mock-treated cells. Western blot analysis of the drug and mock-treated cell lysates with ERK-2 antibody shows the same amount of protein in both lysates, indicating that higher levels of phosphorylated MBP in mock treated cells was not due to an unequal quantity of ERK-2 protein in the lysates. These results suggest that the styryl sulfones of the present invention block the phosphorylating capacity of ERK-2.

The styryl sulfones of the present invention enhance the ability of JNK to phosphorylate c-Jun protein compared to mock-treated cells. Without wishing to be bound by any theory, this result suggests that the styryl sulfones may be acting like pro-inflammatory cytokines or UV light, activating the JNK pathway, which in turn may switch on genes responsible for cell growth inhibition and apoptosis.

### Synthesis of Styryl Sulfones

The compounds of the invention are characterized by cis-trans isomerism resulting from the presence of one or more double bonds. The compounds are named according to the Cahn-Ingold-Prelog system, the IUPAC 1974  
5 Recommendations, Section E: Stereochemistry, in *Nomenclature of Organic Chemistry*, Pergamon, Elmsford, NY, 1979 (the "Blue Book"). See also, March, *Advanced Organic Chemistry*, John Wiley & Sons, Inc., New York, NY, 4th ed., 1992, p. 127-138. Steric relations around a double bond are designated as "Z" or "E".

10 (E)-styryl and benzyl sulfones are prepared by Knoevenagel condensation of aromatic aldehydes with active methylene molecules such as aryl, benzyl, styryl sulfonyl acetic acids, phenacyl aryl sulfones and sulfonyl diacetic acid. The procedure is described by Reddy *et al.*, *Acta. Chim. Hung.* 115:269 (1984); Reddy *et al.*, *Sulfur Letters* 13:83 (1991); Reddy *et al.*, *Synthesis* 322 (1984); and  
15 Reddy *et al.*, *Sulfur Letters* 7:43 (1987), the entire disclosures of which are incorporated herein by reference. (Z)-benzyl and (Z)-styryl sulfones are synthesized by the nucleophilic addition of aromatic and aliphatic thiols to phenyl acetylene, and subsequent oxidation of the product with 30% hydrogen peroxide.

### Preparation of Benzylsulfonyl and Arylsulfonyl Acetic Acids

20 Aryl and benzylsulfonyl acetic acids are the starting compounds for the synthesis of (E)-styryl aryl and (E)-styryl benzyl sulfones. Arylsulfonyl acetic acids may be prepared by the condensation of sodium aryl sulfinic acid with chloroacetic acid at alkaline pH. An alternate method for the synthesis of same compounds involves oxidizing the products obtained by the condensation of sodium arylthiolate  
25 with chloroacetic acid.

Benzylsulfonyl acetic acids may be synthesized by 30% hydrogen peroxide oxidation of the condensation products of the condensation of benzyl chlorides with sodium thioglycollate. Alternatively, benzylsulfonyl acetic acids may be synthesized by 30% hydrogen peroxide oxidation of the products of the  
30 condensation of sodium salts of benzyl thiols with chloroacetic acids.

- 10 -

Synthesis of (E)-Styryl Aryl and (E)-Benzyl Sulfones

To prepare the (E)-styryl benzyl and (E)-styryl benzyl sulfones, a mixture of the appropriate sulfonylacetic acid (e.g., 10 mmol), an aromatic aldehyde (e.g., 10 mmol) and a catalytic amount of benzylamine in acetic acid (e.g., 15 ml) is refluxed for 2-3 hours. After cooling, dry ether is added and the reaction mixture is refrigerated overnight. The ethereal solution is washed successively with a saturated solution of sodium hydrogen carbonate, sodium bisulfite, dilute hydrochloric acid and finally with water. Evaporation of the sodium sulfate dried ethereal solution gives solid products of (E)-styryl aryl or benzyl sulfones which may be recrystallized with 2-propanol or 95% ethanol.

Synthesis of (Z)-Styryl Aryl and (Z)-Styryl Benzyl Sulfones

(Z)-Styryl aryl and (Z)-styryl benzyl sulfones may be prepared by the addition of sodium arylthiolate or benzylthiolate prepared from appropriate thiol (e.g., 10 mmol) and sodium hydroxide (e.g., 20 mmol) to freshly distilled phenylacetylene in methanol. The mixture is refluxed for 24 hours and poured onto crushed ice. The (Z)-styryl aryl and (Z)-styryl benzyl sulfides are oxidized with 30% hydrogen peroxide to provide (Z)-styryl aryl and (Z)-styryl benzyl sulfones, respectively.

Synthesis of (E),(E)- and (E),(Z)-bis(styryl) Sulfones

(E),(E)-bis(styryl) sulfones may be prepared by the condensation of sulfonyl diacetic acid with aromatic aldehydes in the presence of benzylamine as catalyst. The reaction mixture is refluxed for 2 hours in glacial acetic acid. After cooling, absolute ether is added to the reaction mixture, which is washed successively with saturated solution of sodium bicarbonate, sodium bisulfite, dilute hydrochloric acid and water. Evaporation of the dried ethereal layer yields (E),(E)-bis(styryl) sulfones.

(Z),(E)-bis(styryl) sulfones may be prepared by mixing a solution of (Z)-styrylsulfonyl acetic acid in glacial acetic acid with araldehyde and benzylamine. The solution is boiled for 3 hours. The reaction mixture is cooled and dry ether is added. Any product separated is filtered. The filtrate is diluted with more ether and

- 11 -

washed with saturated solution of sodium hydrogen carbonate, sodium bisulfite, dilute hydrochloric acid and water. The ether layer is separated, dried and evaporated to give (Z),(E)-bis(styryl) sulfones.

### **Therapeutic Administration**

5                   The styryl sulfones of the invention may be administered in the form of a pharmaceutical composition, in combination with a pharmaceutically acceptable carrier. The active ingredient in such formulations may comprise from 0.1 to 99.99 weight percent. By "pharmaceutically acceptable carrier" is meant any carrier, diluent or excipient which is compatible with the other ingredients of the formulation and to deleterious to the recipient.

10                   The compounds of the invention may be administered to individuals (mammals, including animals and humans) afflicted with breast or prostate cancer. The compounds may be administered by any route, including oral and parenteral administration. Parenteral administration includes, for example, intravenous, 15 intramuscular, intraarterial, intraperitoneal, intranasal, rectal, or subcutaneous administration. The active agent is preferably administered with a pharmaceutically acceptable carrier selected on the basis of the selected route of administration and standard pharmaceutical practice.

20                   The active agent may be formulated into dosage forms according to standard practices in the field of pharmaceutical preparations. See Gennaro Alphonso, ed., *Remington's Pharmaceutical Sciences*, 18th Ed., (1990) Mack Publishing Co., Easton, PA. Suitable dosage forms may comprise, for example, tablets, capsules, solutions, parenteral solutions, troches, suppositories, or suspensions.

25                   For parenteral administration, the active agent may be mixed with a suitable carrier or diluent such as water, an oil, saline solution, aqueous dextrose (glucose) and related sugar solutions, or a glycol such as propylene glycol or polyethylene glycol. Solutions for parenteral administration preferably contain a water soluble salt of the active agent. Stabilizing agents, antioxidizing agents and preservatives may also be added. Suitable antioxidizing agents include sulfite, 30 ascorbic acid, citric acid and its salts, and sodium EDTA. Suitable preservatives include benzalkonium chloride, methyl- or propyl-paraben, and chlorbutanol.

- 12 -

For oral administration, the active agent may be combined with one or more solid inactive ingredients for the preparation of tablets, capsules, or other suitable oral dosage forms. For example, the active agent may be combined with carboxymethylcellulose calcium, magnesium stearate, mannitol and starch, and then  
5      formed into tablets by conventional tableting methods.

The specific dose of compound according to the invention to obtain therapeutic benefit will, of course, be determined by the particular circumstances of the individual patient including, the size, weight, age and sex of the patient, the nature and stage of the disease, the aggressiveness of the disease, and the route of  
10      administration. For example, a daily dosage of from about 0.05 to about 50 mg/kg/day may be utilized. Higher or lower doses are also contemplated.

The practice of the invention is illustrated by the following non-limiting examples.

#### Procedure 1

##### 15      **General procedure for Synthesis of Styryl and Benzyl Arylsulfones**

To a solution of (8 g, 0.2 mol) sodium hydroxide in methanol (200 ml), appropriate thiophenol or benzyl mercaptan (0.1 mol) is added slowly. Then chloroacetic acid (0.1 mol) is added in portions and the reaction mixture is refluxed for 2-3 hours. The cooled contents are poured onto crushed ice and neutralized  
20      with dilute hydrochloric acid (200 ml). The resulting aryl and benzylthioacetic acids (0.1 mol) are subjected to oxidation with 30% hydrogen peroxide (0.12 mol) in glacial acetic acid (25 ml) by refluxing for 1-2 hours. The contents are cooled and poured onto crushed ice. The separated solid is recrystallized from hot water to give pure aryl and benzylsulfonyl acetic acids.

25      A mixture of the appropriate aryl or benzylsulfonyl acetic acid (0.001 mol), an aromatic aldehyde (0.001 mol) and benzylamine (1 ml) in glacial acetic acid (15 ml) is refluxed for 2-3 hours. The contents are cooled and treated with dry ether (50 ml). Any product separated is collected by filtration. The filtrate is diluted with more ether and washed successively with a saturated solution  
30      of sodium bicarbonate (20 ml), sodium bisulfite (20 ml), dilute hydrochloric acid

- 13 -

(20 ml) and finally with water (35 ml). Evaporation of the dried ethereal layer yields a solid in many cases. However, in some cases a syrupy material separates and is solidified on treatment with 2-propanol. The purity of the compounds is checked by TLC (silica gel BDH, hexane/ethyl acetate 3:1).

5

## Procedure 2

### General procedure for Synthesis of (E)(E)- and (E)(Z)-bis(Styryl) Sulfones

To freshly distilled phenyl acetylene (51.07 g, 0.5 mol) is added sodium thioglycollate prepared from thioglycolic acid (46 g, 0.5 mol) and sodium hydroxide (40 g, 1 mol) in methanol (250 ml). The mixture is refluxed for 24  
10 hours and poured onto crushed ice (500 ml) after cooling. The styrylthioacetic acid, formed after neutralization with dilute hydrochloric acid (250 ml), is filtered and dried; yield 88 g (90%); m.p. 84-86°C.

The styrylthioacetic acid is then oxidized to styrylsulfonylacetic acid as follows. A mixture of styrylthioacetic acid (5 g, 25 mmol) in glacial acetic acid  
15 (35 ml) and 30% hydrogen peroxide (15 ml) is heated under reflux for 60 minutes and the mixture is poured onto crushed ice (200 ml) after cooling. The compound separated is filtered and recrystallized from hot water to give white crystalline flakes of (Z)-styrylsulfonylacetic acid; yield 2.4 g (41%); m.p. 150-51°C.

A solution of (Z)-styrylsulfonylacetic acid (2.263 g, 10 mmol) in  
20 glacial acetic acid (6 ml) is mixed with an aromatic aldehyde (10 mmol) and benzylamine (0.2 ml) and refluxed for 3 hours. The reaction mixture is cooled, treated with dry ether (50 ml), and any product separated is collected by filtration. The filtrate is diluted with more ether and washed successively with a saturated solution of sodium hydrogen carbonate (15 ml), sodium bisulfite (15 ml), dilute  
25 hydrochloric acid (20 ml) and finally with water (30 ml). Evaporation of the dried ethereal layer yields (E)(Z)-bis(styryl)sulfones.

(E),(E)-bis(styryl)sulfones are prepared following the same procedure as described above with exception that sulfonyldiacetic acid is used in place of (Z)-styrylsulfonylacetic acid, and twice the amount of aromatic aldehyde  
30 (20 mmol) is used.



### Procedure 3

#### General Procedure for the Synthesis of 2-(Arylsulfonyl)-1-phenyl-3-aryl-2-propen-1-ones

These compounds are synthesized by two methods which employ  
5 different reaction conditions, solvents and catalysts.

**Method 1:** Phenacyl aryl sulfones are made by refluxing  $\alpha$ -bromoacetophenones (0.05 mol) and sodium arylsulfonates (0.05 mol) in absolute ethanol (200 ml) for 6-8 hours. The product which separates on cooling is filtered and washed several times with water to remove sodium bromide. The product is  
10 then recrystallized from ethanol: phenacyl-phenyl sulfone. m.p. 90-91°C; phenacyl-p-fluorophenyl sulfone. m.p. 148-149°C; phenacyl-p-bromophenyl sulfone, m.p. 121-122°C; phenacyl-p-methoxyphenyl sulfone. m.p. 104-105°C; p-nitrophenacyl-phenyl sulfone, m.p. 136-137°C.

A solution of phenacyl aryl sulfone (0.01 mol) in acetic acid (10 ml)  
15 is mixed with an araldehyde (0.01 mol) and benzylamine (0.02 ml) and refluxed for 3 hours. The solution is cooled and dry ether (50 ml) is added. The ethereal solution is washed successively with dilute hydrochloric acid, aqueous 10% NaOH, saturated NaHSO<sub>3</sub> solution and water. Evaporation of the dried ethereal layer gives a solid product which is purified by recrystallization.

**Method 2:** Dry tetrahydrofuran (200 ml) is taken in a 500 ml conical  
20 flask flushed with nitrogen. To this, a solution of titanium (IV) chloride ( 11 ml, 0.01 mol) in absolute carbon tetrachloride is added dropwise with continuous stirring. The contents of the flask are maintained at -20°C throughout the course of the addition. A mixture of phenacyl aryl sulfone (0.01 mol) and aromatic  
25 aldehyde (0.01 mol) is added to the reaction mixture and pyridine (4 ml, 0.04 mol) in tetrahydrofuran (8 ml) is added slowly over a period of 1 hour. The contents are stirred for 10-12 hours, treated with water (50 ml) and then ether (50 ml) is added. The ethereal layer is separated and washed with 15 ml of saturated solutions of 10%

- 15 -

sodium hydroxide, sodium bisulfite and brine. The evaporation of the dried ethereal layer yields 2-(arylsulfonyl)-1-phenyl-3-aryl-2 propen-1-ones.

#### Example 1

##### E-styryl phenyl sulfone

5                   A solution of phenyl sulfonylacetic acid (0.01 mol) and benzaldehyde (0.01 mol) was subjected to the Procedure 1. The title compound was obtained in 68-72% yield.

#### Example 2

##### E-4-chlorostyryl phenyl sulfone

10                   A solution of phenyl sulfonylacetic acid (0.01 mol) and 4-chlorobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound was obtained in 78-80% yield.

#### Example 3

##### E-2,4-dichlorostyryl phenyl sulfone

15                   A solution of phenyl sulfonylacetic acid (0.01 mol) and 2,4-dichlorobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound was obtained in 60-65% yield.

#### Example 4

##### E-4-bromostyryl phenyl sulfone

20                   A solution of phenyl sulfonylacetic acid (0.01 mol) and 4-bromobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound was obtained in 78-80% yield.

- 16 -

**Example 5****E-4-chlorostyryl 4-chlorophenyl sulfone**

A solution of 4-chlorophenyl sulfonylacetic acid (0.01 mol) and 4-chlorobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound  
5 was obtained in 70-72 % yield.

**Example 6****E-4-methylstyryl 4-chlorophenyl sulfone**

A solution of 4-chlorophenyl sulfonylacetic acid (0.01 mol) and 4-methylbenzaldehyde (0.01 mol) was subjected to Procedure 1. The title  
10 compound was obtained in 60-64 % yield.

**Example 7****E-4-methoxystyryl 4-chlorophenyl sulfone**

A solution of 4-chlorophenyl sulfonylacetic acid (0.01 mol) and 4-methoxybenzaldehyde (0.01 mol) was subjected to Procedure 1. The title  
15 compound was obtained in 68-70 % yield.

**Example 8****E-4-bromostyryl 4-chlorophenyl sulfone**

A solution of 4-chlorophenyl sulfonylacetic acid (0.01 mol) and 4-bromobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound  
20 was obtained in 80 % yield.

**Example 9****E-2-chlorostyryl benzyl sulfone**

A solution of benzyl sulfonylacetic acid (0.01 mol) and 2-chlorobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title  
25 compound was obtained in 72 % yield.

- 17 -

**Example 10****E-4-chlorostyryl benzyl sulfone**

A solution of benzyl sulfonylacetic acid (0.01 mol) and 4-chlorobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound was obtained in 78% yield.

**Example 11****E-4-fluorostyryl 4-chlorobenzyl sulfone**

A solution of 4-chlorobenzyl sulfonylacetic acid (0.01 mol) and 4-fluorobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound was obtained in 72% yield.

**Example 12****E-4-chlorostyryl 4-chlorobenzyl sulfone**

A solution of 4-chlorobenzyl sulfonylacetic acid (0.01 mol) and 4-chlorobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound was obtained in 80% yield.

**Example 13****E-4-fluorostyryl 4-fluorobenzyl sulfone**

A solution of 4-fluorobenzyl sulfonylacetic acid (0.01 mol) and 4-fluorobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound was obtained in 73% yield.

**Example 14****E-2,4-difluorostyryl 4-fluorobenzyl sulfone**

A solution of 4-fluorobenzyl sulfonylacetic acid (0.01 mol) and 2,4-difluorobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound was obtained in 68% yield.

- 18 -

**Example 15****E-4-fluorostyryl 4-bromobenzyl sulfone**

A solution of 4-bromobenzyl sulfonylacetic acid (0.01 mol) and 4-fluorobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound was obtained in 82% yield.

**Example 16****E-4-bromostyryl 4-bromobenzyl sulfone**

A solution of 4-bromobenzyl sulfonylacetic acid (0.01 mol) and 4-bromobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound was obtained in 88% yield.

**Example 17****E-4-bromostyryl 4-fluorobenzyl sulfone**

A solution of 4-fluorobenzyl sulfonylacetic acid (0.01 mol) and 4-bromobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound was obtained in 82% yield.

**Example 18****E-4-chlorostyryl 4-bromobenzyl sulfone**

A solution of 4-bromobenzylsulfonyl acetic acid (0.01 mol) and 4-chlorobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound was obtained in 88% yield.

**Example 19****E-4-bromostyryl 4-chlorobenzyl sulfone**

A solution of 4-chlorobenzylsulfonyl acetic acid (0.01 mol) and 4-bromobenzaldehyde (0.01 mol) was subjected to Procedure 1. The title compound was obtained in 92% yield.

- 19 -

**Example 20****(Z)-styryl-(E)-4-fluorostyryl sulfone**

A solution of (Z)-styryl sulfonylacetic acid (0.01 mol) and 4-4-fluorobenzaldehyde (0.01 mol) was subjected to Procedure 2. The title compound  
5 was obtained in 68% yield.

**Example 21****(Z)-styryl-(E)-4-bromostyryl sulfone**

A solution of (Z)-styryl sulfonylacetic acid (0.01 mol) and 4-bromobenzaldehyde (0.01 mol) was subjected to Procedure 2. The title  
10 compound was obtained in 70% yield.

**Example 22****(Z)-styryl-(E)-4-chlorostyryl sulfone**

A solution of (Z)-styryl sulfonylacetic acid (0.01 mol) and 4-chlorobenzaldehyde (0.01 mol) was subjected to Procedure 2. The title  
15 compound was obtained in 64% yield.

**Example 23****2-[(4-fluorophenyl)sulfonyl]-1-phenyl-3-(4-fluorophenyl)-2-propen-1-one**

A solution of phenacyl-4-fluorophenyl sulfone (0.01 mol) and 4-fluorobenzaldehyde (0.01 mol) was subjected to Method 1 of Procedure 3. The title  
20 compound was obtained in 63% yield.

**Example 24****2-[(4-fluorophenyl-sulfonyl)-1-phenyl-3-(4-fluorophenyl)-2-propen-1-one**

A solution of phenacyl-2-chlorophenyl sulfone (0.01 mol) and 2-fluoro benzaldehyde (0.01 mol) was subjected to Method 1 of Procedure 3. The  
25 title compound was obtained in 58% yield.

- 20 -

**Example 25****2-[(2-chlorophenyl)sulfonyl]-1-phenyl-3-(4-bromophenyl)-2-propen-1-one**

A solution of phenacyl-2-chlorophenyl sulfone (0.01 mol) and 4-bromo benzaldehyde (0.01 mol) was subjected to Method 1 of Procedure 3. The title compound was obtained in 66% yield.

**Example 26****2-[(4-chlorophenyl)sulfonyl]-1-phenyl-3-(4-bromophenyl)-2-propen-1-one**

A solution of phenacyl-4-chlorophenyl sulfone (0.01 mol) and 4-bromo benzaldehyde (0.01 mol) was subjected to Method 1 of Procedure 3. The title compound was obtained in 60% yield.

**Example 27****2-[(2-nitrophenyl)sulfonyl]-1-phenyl-3-(4-bromophenyl)-2-propen-1-one**

A solution of phenacyl-2-nitrophenyl sulfone (0.01 mol) and 4-bromo benzaldehyde (0.01 mol) was subjected to Method 1 of Procedure 3. The title compound was obtained in 56% yield.

**Example 28****Tumor Cell Growth Inhibition by Styryl Sulfones****A. Cells.**

The effect of the styryl sulfones on the growth of normal and tumor cells of breast and prostate was examined utilizing four cell lines. NIH3T3, MCF-7, BT-20 and LnCap. NIH3T3 cells represent normal fibroblasts while LnCap is an androgen-dependent prostate tumor cell line. MCF-7 is an estrogen-responsive breast tumor cell line, while BT-20 is an estrogen-unresponsive breast tumor cell line. MCF-7 and BT-20 were grown in Dulbecco's modified Eagle's medium (DMEM) containing 10% fetal bovine serum supplemented with penicillin and streptomycin. LnCap were cultured in RPMI with 10% fetal bovine serum containing penicillin and streptomycin. NIH3T3 cells were grown in DMEM containing 10% calf serum

- 21 -

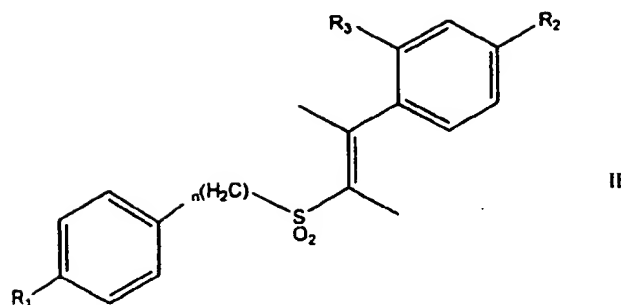
supplemented with penicillin and streptomycin. All cell cultures were maintained at 37°C in a humidified atmosphere of 5% CO<sub>2</sub>.

B. Treatment with Styryl Sulfones and Viability Assay

5 Cells were treated with test compound at 2.5 µM or 5.0 µM concentration and cell viability was determined after 48 hours by the Trypan blue exclusion method. The compounds identified in Tables 1, 2 and 3 inhibited cell growth and induced cell death, to varying degrees. The tables list the percent viable LnCap and MCF-7 cells treated with 5.0 µM compound.

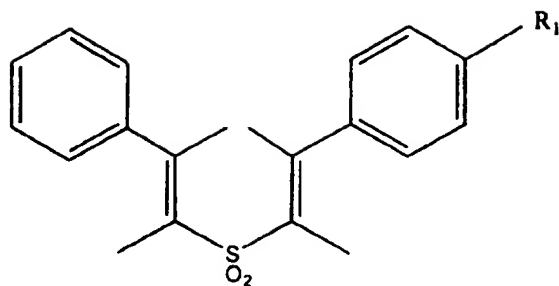


Table I



	Example	n	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>	% viable LnCaP and MCF-7 cells
5	1	0	H	H	H	89
	2	0	H	Cl	H	90
	3	0	H	Cl	Cl	88
	4	0	H	Br	H	68
	5	0	Cl	Cl	H	64
	6	0	Cl	CH <sub>3</sub>	H	92
10	7	0	Cl	OCH <sub>3</sub>	H	90
	8	0	Cl	Br	H	69
	9	1	H	H	Cl	94
	10	1	H	Cl	H	87
15	11	1	Cl	F	H	6
	12	1	Cl	Cl	H	49
	13	1	F	F	H	43
	14	1	F	F	F	56
	15	1	Br	F	H	7
	16	1	Br	Br	H	51
20	17	1	F	Br	H	42
	18	1	Br	Cl	H	7
	19	1	Cl	Br	H	20

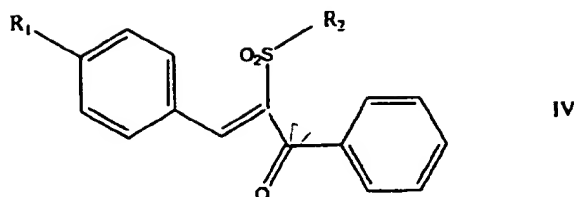
Table 2



III

Example	R <sub>1</sub>	% viable LnCaP and MCF-7 cells
20	F	76
21	Br	68
22	Cl	72

Table 3



Example	R <sub>1</sub>	R <sub>2</sub>	% viable LnCaP and MCF-7 cells
23	F	4-fluorophenyl	76
24	F	2-chlorophenyl	64
5 25	Br	2-chlorophenyl	72
26	Br	4-chlorophenyl	58
27	Br	2-nitro	74

Five of the more active compounds which exhibited the highest activity were designated as FRI-2 (E-2,4-difluorostyryl-4-fluorobenzyl sulfone), FRI-6 (E-4-fluorostyryl 4-bromobenzyl sulfone), FRI-7 (E-4-bromostyryl 4-fluorobenzyl sulfone), FRI-20 (E-4-fluorostyryl 4-chlorobenzyl sulfone) and FRI-22 (E-4-chlorostyryl 4-chlorobenzyl sulfone). These compounds were found to substantially inhibit the growth and induce the death of LnCaP, BT-20 and MCF-7 cells at 2.5 mM (Figure 1A) and 5.0 mM (Figure 1B), after 48 hours of treatment with the compounds. Under identical conditions, more than 80% of NIH3T3 cells were viable after 48 hours incubation (Figure 1A and 1B). E-4-chlorostyryl 4-bromobenzyl sulfone and E-4-bromostyryl 4-chlorobenzyl sulfone were also highly active.

#### C. Dose-Dependency Assay

The dose dependency of the styryl sulfone was established by treating the cells with FRI-20, one of the five most active compounds. NIH3T3, MCF-7, BT-20 and

- 25 -

LnCaP cells were treated with FRI-20 dissolved in DMSO to concentrations of 250 nM, 500 nM, 1  $\mu$ M, 2.5  $\mu$ M and 5  $\mu$ M and examined for their proliferation and viability after 48 hours (Figure 2A). The percentage of living cells was determined by Trypan blue exclusion. The control cells were treated with DMSO to determine the effect of solvent on cells. At a concentration of 250 nM, there was about 10% cell death in MCF-7, BT-20 and LnCaP cells and about 15-20% inhibition in cell division compared to untreated cells after 48 hours. There was about 30-50% inhibition in cell proliferation and 25-30% cell death in LnCap. BT-20 and MCF-7 at 500 nM concentration. Under these conditions, only 2-3% of NIH3T3 cells were non-viable at both the concentrations. The LnCaP, BT-20 and MCF-7 cell growth was greatly inhibited by 1  $\mu$ M concentration of FRI-20 with concomitant loss of cell viability. After 48 hours incubation, 60-75% of the LnCaP, BT-20 and MCF-7 cells were dead at 2.5 mM FRI-20 concentration, whereas more than 90% of NIH3T3 cells were viable (Figure 2A). The LnCap. BT-20 and MCF-7 cells treated with 5  $\mu$ M FRI-20 (Figure 2A) showed nearly 90% cell death. NIH3T3 showed little or no alteration in their ability to grow and maintain >80% viability in the presence of FRI-2. -6. -7, -20 or -22, at 5  $\mu$ M concentration.

#### D. Time Course Assay

The time course of the activity of FRI-20 was demonstrated as follows. NIH/3T3, MCF-7, BT-20 and LnCaP were treated with FRI-20 at 2.5  $\mu$ M and the number of viable cells was determined at 12, 24, 48 and 72 hours by Trypan blue exclusion. The mean of three independent experiments is shown in Fig. 2B. The time course study revealed that more than 95% of MCF-7, LnCaP, and BT-20 cells were dead after 72 hours of treatment with FRI-20 at 2.5  $\mu$ M (Figure 2B).

25

### Example 29

#### Tumor Cell Growth Inhibition by FRI-20

##### A. Cells.

The effect of FR-20 on the growth of normal and tumor cells of breast and prostate was examine utilizing nine cell lines: NIH/3T3 and HFL (normal fibroblast

- 26 -

cell lines); MCF-7 and 361 (estrogen-receptor negative breast tumor cell lines); BT-20, 435 and SKBR-3 (estrogen-receptor positive breast tumor cell lines); LnCaP (androgen sensitive prostate tumor cell line); PC-3 and DU-145 (androgen insensitive prostate tumor cell line).

5 B. Treatment with FRI-20 and Viability Assay.

The cells were grown as in Example 22, A. FR-20 was dissolved in DMSO and added to the cells at 2.5  $\mu$ M and 5.0  $\mu$ M concentration. To control cells, DMSO was added equivalent to the volume of solvent (DMSO) present at the highest concentration of the compound. The activity of the compound was evaluated after  
10 48 hours by Trypan blue exclusion. NIH3T3 and HFL cells were found to maintain a percent viability of 85-90% at 2.5 and 5.0  $\mu$ M concentration. Of the seven breast tumor cell lines treated with FRI-20 compound, MCF-7, HTB126, T470 and 435 cells showed very high mortality with less than 25% and 10% viability at 2.5 and 5.0  $\mu$ M concentrations of the drug (Figure 3A). Nearly 50% of SKBR-3 and BT-20 cells  
15 were dead at 2.5  $\mu$ M and 75% at 5.0  $\mu$ M concentration of the compound. The 361 breast tumor cell line, on the other hand showed considerable resistance to FRI-20 with 50-75% of cells being viable at 2.5 and 5.0  $\mu$ M concentration. FRI-20 had profound effect on the viability of androgen-dependent LnCaP prostate tumor cell line when compared to androgen-independent DU-145 and PC-3 prostate cell lines.  
20 At 2.5 mM FRI-20, 80% of LnCaP, 40% of PC-3 and 20% of DU-145 cells were killed. At 5.0 mM FRI-20, 72% of LnCaP, 47% of PC-3 and 40% of DU-145 were killed (Figure 3B).

Example 30

Effect of FRI-20 on Cell Cycle Regulation

25 The androgen-dependent prostate tumor cell line LnCaP was grown as in Ex. 22, A, and treated with 2.0  $\mu$ M FRI-20 dissolved in DMSO or with equivalent amounts (10 ml) of DMSO alone. Cells were harvested 6, 12, 24, and 48 hours following treatment and stained with propidium iodide and subjected to flow cytometry (FACS) for analysis of DNA content. As shown in Fig. 4, the addition of  
30 FR-20 to the culture medium results in the accumulation of cells in the G2/M phase

- 27 -

of the cell cycle and as the cells exit this phase of the cell cycle, they appeared to undergo apoptosis. Cells treated with DMSO alone failed to exhibit such an arrest in the G2/M phase of the cell cycle, suggesting that the effects seen are associated with FRI-20 addition. Treatment of the normal cell lines NIH3T3 or HFL with FRI-20 failed to produce a similar effect on cell cycle progression. NIH3T3 and HFL exhibited normal cell cycle progression in the presence and absence of drug.

### Example 31

#### Effect of FRI-20 on MPK Pathway

##### A. Immune Complex ERK-2 Assay.

To examine the effects of FRI-20 on the MAPK pathway, NIH3T3, LnCaP and MCF-7 cells were incubated with FRI-20 at a concentration 2.5 mM for 48 hours. Following incubation of cells in the presence and absence of FRI-20, the cells were lysed using ERK lysis buffer containing 20 mM HEPES (pH 7.4), 50 mM  $\beta$ -glycerophosphate, 0.5% Triton X-100, 2 mM  $MgCl_2$ , 1 mM EGTA, 1 mM dithiothreitol, 2  $\mu$ g/ml leupeptin, 2  $\mu$ g/ml aprotinin, 100  $\mu$ M phenylmethylsulfonyl fluoride, and 1 mM benzamidine. ERK-2 in 100 mg of cell lysate was immunoprecipitated by incubating lysate protein with 1 mg of ERK-2 polyclonal antibody (antibody sc-154 to ERK2 is from Santa Cruz Biotechnology, Inc.) for one hour followed by an additional incubation of 20  $\mu$ l of protein A-Sepharose (Pharmacia) for one hour. The immune complex-bound protein A-Sepharose beads were washed twice with lysis buffer and twice with ERK/MAPK buffer containing 20 mM HEPES (pH 7.4), 50 mM  $\beta$ -glycerophosphate, 10 mM  $MgCl_2$ , 1 mM EGTA, 1 mM dithiothreitol, and 100 mM  $Na_3VO_4$ .

The immunoprecipitates were then tested for MAP kinase activity by an *in vitro* assay which utilizes myelin basic proteins (MBP) as a substrate for ERK-2 in the presence of [ $\gamma$ - $^{32}P$ ] ATP. Accordingly, the beads were resuspended in 40  $\mu$ l of MAPK buffer containing 100  $\mu$ M [ $\gamma$ - $^{32}P$ ] ATP (5000 cpm/pmol), and the kinase assay was carried out for 20 minutes at 30°C using 5  $\mu$ g of MBP as substrate. The reaction was stopped by the addition of Laemmli's buffer followed by the boiling of the samples for 3 minutes. The proteins were resolved on 12% SDS-PAGE; the gel

- 28 -

was dried, and an autoradiogram was developed. The results show that both drug-treated and untreated cells exhibit similar levels of intracellular ERK-2, but the biochemical activity of ERK-2, as judged by its ability to phosphorylate MBP, was considerably diminished in drug-treated cells compared to cells treated with DMSO alone. In prostate tumor cells, FRI-20 reduced the phosphorylation status of MBP by more than 80% compared to mock-treated cells (Figure 5).

B. Western Blot Analysis.

Cell lysates of FRI-20-treated cells were prepared for Western Blot analysis as follows. NIH3T3, LnCaP or MCF-7 cells were seeded at a density of  $2 \times 10^5$  cells/per well in a six-well plate and allowed to grow for 24 hours. Fresh medium was added to each well 2 hours before treatment with FRI-20. The compound was dissolved in DMSO to make a 2 mM stock solution and added to the medium (2 ml) to obtain a final concentration of 2.5 and 5.0  $\mu$ M. After 48 hours at 37°C, the cells were washed twice with 10 ml of ice cold phosphate-buffered saline and harvested in 400  $\mu$ l of lysis buffer containing 25 mM HEPES (pH 7.6), 0.1% Triton X-100, 300 mM NaCl, 1.5 mM  $MgCl_2$ , 20 mM  $\beta$ -glycerophosphate, 100  $\mu$ M  $Na_3VO_4$ , 0.2 mM EDTA, 0.5 mM dithiothreitol, 2  $\mu$ g/ml aprotinin, 2  $\mu$ g/ml leupeptin, 100  $\mu$ M phenylmethylsulfonyl chloride and 1 mM benzamidine. The cell lysates were kept on ice for 30 minutes and centrifuged for 10 minutes in a microcentrifuge (16000 X g). The cell lysates were separated from the debris and normalized for protein content.

Western Blot analysis was carried out on the drug- and mock-treated cell lysates with ERK-2 antibody. Equal amounts of total protein (100  $\mu$ g) were loaded in each lane of a SDS-PAGE gel (10-12%) and transferred to Immobilon-P (Millipore, USA). Following transfer, membranes were blocked in 3% milk, then probed with ERK-2 and JNK-1 rabbit polyclonal antibodies (Santa Cruz Biotechnology Inc., Santa Cruz, CA) and then probed with horseradish peroxidase linked donkey anti-rabbit Ig secondary antibody (Amersham) (1:10000 dilution). The antibody was detected using the ECL Western blotting analysis kit (Amersham) following the manufacturer's instructions. The western blot analysis of the drug- and

- 29 -

mock-treated cell lysates with ERK-2 antibody showed the same amount of protein in both lysates (Figure 6), indicating that higher levels of MBP phosphorylation in mock-treated cells was not due to an unequal quantity of ERK-2 protein in lysates. These results suggest that FRI-20 blocks the phosphorylating capability of ERK-2.

5

### **Example 32**

#### **Effect of FRI-20 on Stress-Activated Protein Activity**

To further establish if the activity of stress activated protein kinases (SAPKs), of which JNK is a member, is compromised in the presence of FRI-20, cells (NIH3T3, MCF-7 or LnCaP) were treated with FRI-20 dissolved in DMSO or with DMSO alone. Forty-eight hours later, the cells were lysed with kinase buffer and the lysates used for estimation of the amount of JNK present in each lysate by western blot analysis using JNK polyclonal antibody. The biochemical activity of the JNK present in the FRI-20-treated and mock-treated cell lysates was also determined by immunoprecipitation of JNK followed by incubation with GST-c-Jun protein as a substrate for JNK in the presence of [ $\gamma$ - $^{32}$ P]ATP.

Accordingly, JNK-1 in 100 mg of cell extracts was immunoprecipitated by incubating the lysate with 1 mg of JNK-1 polyclonal antibody (sc from Santa Cruz Biotechnology) for one hour followed by an additional incubation with 20  $\mu$ l of protein A-Sepharose (Pharmacia) for one hour. The beads were washed twice with JNK lysis buffer (as described above) followed by two washes with JNK reaction buffer. The beads were resuspended in 40  $\mu$ l of JNK buffer containing 20 mM [ $\gamma$ - $^{32}$ P] ATP (5000 cpm/pmol), and the kinase reaction was carried out for 20 minutes at 30°C using 3  $\mu$ g of purified GST-c-Jun(1-79) as substrate. The reaction was stopped, and the radioactivity in the phosphorylated GST-c-Jun protein was quantitated. The results show that the FRI-20 treatment enhanced the ability of JNK to phosphorylate recombinant GST-c-Jun protein by 60-80% compared to mock-treated cells (Figure 7).

JNK has been shown to be activated by treatment of cells with UV radiation, pro-inflammatory cytokines and environmental stress (Derijard *et al.*, *Cell* 1025 (1994)). The activated JNK binds to the amino terminus of c-jun and increases its transcriptional activity by phosphorylating at ser63 and ser73 (Adler *et al.*, *Proc.*



- 30 -

*Natl. Acad. Sci. USA* **89**:5341 (1992); Kwok *et al.*, *Nature* **370**:223 (1994)). Without wishing to be bound by any theory, the results demonstrated herein suggest that FRI-20 may act like a pro-inflammatory cytokine or UV light in activating the JNK pathway, which in turn may switch on genes responsible for cell growth inhibition and apoptosis.

### Example 33

#### **Comparison of FRI-20 and Cisplatin**

##### Anti-Tumor Activities

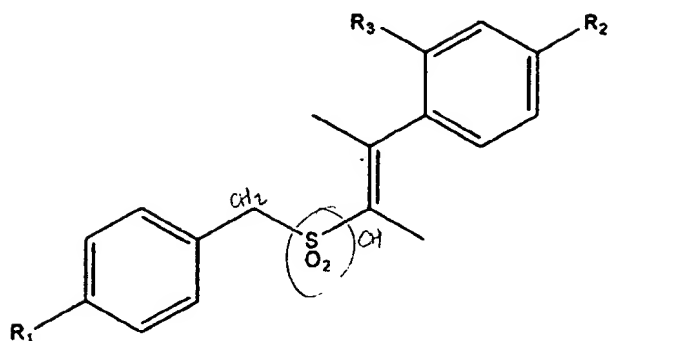
The killing effect of FR-20 on androgen-sensitive (LnCaP) and androgen insensitive (DU145) prostate tumor cells was compared to the effect of cisplatin (cis-diamminedichloroplatinum II), a widely used anti-prostate cancer agent. The cells were grown as in Example 26. FRI-20 or cisplatin was dissolved in DMSO and added to the cells at various concentrations. Viability was determined after 72 hours by the Trypan blue exclusion method. The concentration of FRI-20 required to completely kill LnCaP and DU145 cells was 2.5  $\mu$ M and 5.0  $\mu$ M, respectively. Under identical conditions, complete killing of LnCaP and DU145 cells by cisplatin required 25  $\mu$ M and 15  $\mu$ M concentrations, respectively. Thus, FRI-20 is at least tenfold more active than cisplatin in killing both hormone-dependent and hormone-independent prostate tumor cells.

All references cited with respect to synthetic, preparative and analytical procedures are incorporated herein by reference.

The present invention may be embodied in other specific forms without departing from the spirit or essential attributes thereof and, accordingly, reference should be made to the appended claims, rather than to the foregoing specification, as indication the scope of the invention.

**CLAIMS**

1. A compound of the formula I:



5 wherein

$R_1$  and  $R_2$  are independently selected from the group consisting of chlorine, fluorine and bromine; and

$R_3$  is selected from the group consisting of hydrogen and fluorine:

$R_1$  and  $R_2$  may not both be chlorine when  $R_3$  is hydrogen; and

10  $R_1$  may not be chlorine when  $R_2$  is fluorine and  $R_3$  is hydrogen in the same compound.

2. A compound according to claim 1 wherein the compound is E-4-fluorostyryl 4-fluorobenzyl sulfone.

3. A compound according to claim 1 wherein the compound is E-2,4-  
15 difluorostyryl 4-fluorobenzyl sulfone.

4. A compound according to claim 1 wherein the compound is E-4-fluorostyryl 4-bromobenzyl sulfone.

5. A compound according to claim 1 wherein the compound is E-4-bromostyryl 4-bromobenzyl sulfone.

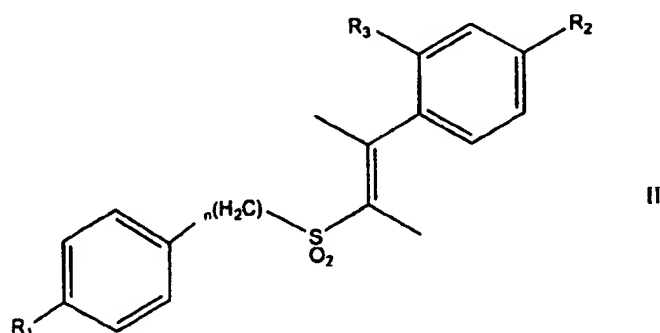
- 32 -

6. A compound according to claim 1 wherein the compound is E-4-chlorostyryl 4-bromobenzyl sulfone.

7. A compound according to claim 1 wherein the compound is E-4-bromostyryl 4-chlorobenzyl sulfone.

5 8. A compound according to claim 1 wherein the compound is E-4-bromostyryl 4-fluorobenzyl sulfone.

9. A pharmaceutical composition comprising a pharmaceutically acceptable carrier and a compound of the formula II



wherein

10 n is zero or one;  
 $R_1$  is selected from the group consisting of hydrogen, chlorine, fluorine and bromine;

$R_2$  is selected from the group consisting of hydrogen, chlorine, fluorine, bromine, methyl and methoxy; and

15  $R_3$  is selected from the group consisting of hydrogen, chlorine and fluorine;

provided,

$R_2$  may not be methyl or methoxy when  $R_1$  and  $R_3$  are both hydrogen and n is zero or one; and

20  $R_1$ ,  $R_2$  and  $R_3$  may not all be hydrogen when n is one.

- 33 -

10. A composition according to claim 9 wherein the compound is E-4-bromostyryl phenyl sulfone.

11. A composition according to claim 9 wherein the compound is E-2,4-difluorostyryl 4-fluorobenzyl sulfone.

5 12. A composition according to claim 9, wherein  $R_3$  is hydrogen, and  $R_1$  and  $R_2$  are independently selected from the group consisting of chlorine, fluorine and bromine.

13. A composition according to claim 12 wherein the compound is E-4-chlorostyryl 4-chlorophenyl sulfone.

10 14. A composition according to claim 12 wherein the compound is E-4-bromostyryl 4-chlorophenyl sulfone.

15. A composition according to claim 12 wherein the compound is E-4-fluorostyryl 4-chlorobenzyl sulfone.

15 16. A composition according to claim 12 wherein the compound is E-4-chlorostyryl 4-chlorobenzyl sulfone.

17. A composition according to claim 12 wherein the compound is E-4-fluorostyryl 4-fluorobenzyl sulfone.

18. A composition according to claim 12 wherein the compound is E-4-fluorostyryl 4-bromobenzyl sulfone.

20 19. A composition according to claim 12 wherein the compound is E-4-bromostyryl 4-bromobenzyl sulfone.

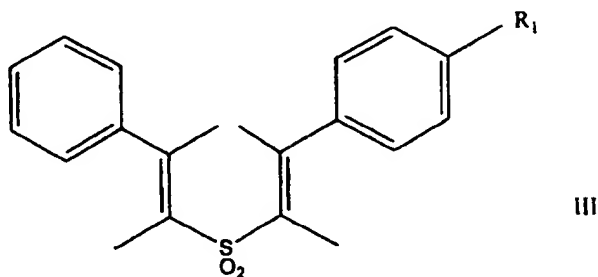
- 34 -

20. A composition according to claim 12 wherein the compound is E-4-bromostyryl 4-fluorobenzyl sulfone.

21. A composition according to claim 12 wherein the compound is E-4-chlorostyryl 4-bromobenzyl sulfone.

5 22. A composition according to claim 12 wherein the compound is E-4-bromostyryl 4-chlorobenzyl sulfone.

23. A pharmaceutical composition comprising a pharmaceutically acceptable carrier and a compound of the formula III

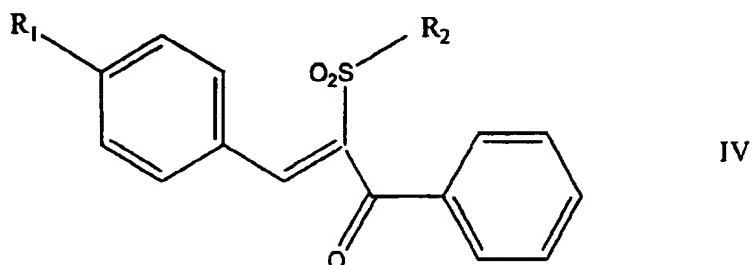


wherein

10

$R_1$  is selected from the group consisting of hydrogen, chlorine, fluorine and bromine.

24. A pharmaceutical composition comprising a pharmaceutically acceptable carrier and the compound of formula IV



wherein

5         $R_1$  is selected from the group consisting of fluorine and bromine, and  $R_2$  is selected from the group consisting of 2-chlorophenyl, 4-chlorophenyl, 4-fluorophenyl and 4-nitro.

25. A method of treating an individual for breast or prostate cancer comprising administering to said individual an effective amount of a pharmaceutical composition according to claim 9.

10        26. A method of inhibiting growth of breast or prostate tumor cells in an individual afflicted with breast or prostate cancer comprising administering to said individual a pharmaceutical composition according to claim 9.

✓  
15        27. A method of inducing apoptosis of breast or prostate tumor cells in an individual afflicted with breast or prostate cancer comprising administering to said individual a pharmaceutical composition according to claim 9.

28. A method of treating an individual for breast or prostate cancer comprising administering to said individual an effective amount of a pharmaceutical composition according to claim 23 or 24.

- 36 -

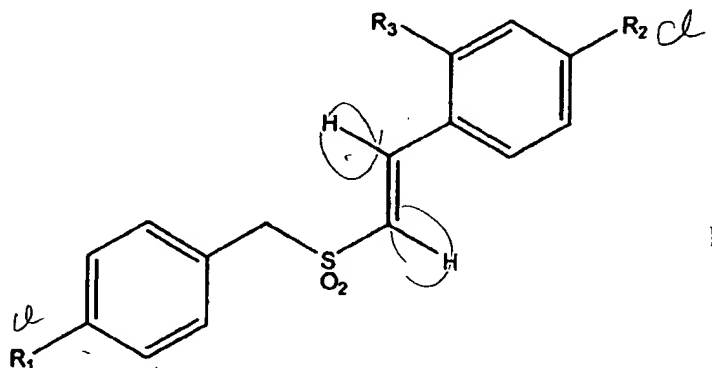
29. A method of inhibiting growth of breast or prostate tumor cells in an individual afflicted with breast or prostate cancer comprising administering to said individual a pharmaceutical composition according to claim 23 or 24.

5 30. A method of inducing apoptosis of breast or prostate tumor cells in an individual afflicted with breast or prostate cancer comprising administering to said individual a pharmaceutical composition according to claim 23 or 24.

## AMENDED CLAIMS

[received by the International Bureau on 23 February 1999 (23.02.99);  
original claims 1, 9, 23-30 amended; new claims 31-36 added; remaining claims unchanged  
(5 pages)]

1. A compound of the formula I:



5 wherein

$R_1$  and  $R_2$  are independently selected from the group consisting of chlorine, fluorine and bromine; and

$R_3$  is selected from the group consisting of hydrogen and fluorine;

$R_1$  and  $R_2$  may not both be chlorine when  $R_3$  is hydrogen; and

10  $R_1$  may not be chlorine when  $R_2$  is fluorine and  $R_3$  is hydrogen in the same compound.

2. A compound according to claim 1 wherein the compound is E-4-fluorostyryl 4-fluorobenzyl sulfone.

3. A compound according to claim 1 wherein the compound is E-2,4-  
15 difluorostyryl 4-fluorobenzyl sulfone.

4. A compound according to claim 1 wherein the compound is E-4-fluorostyryl 4-bromobenzyl sulfone.

5. A compound according to claim 1 wherein the compound is E-4-bromostyryl 4-bromobenzyl sulfone.

AMENDED SHEET (ARTICLE 19)

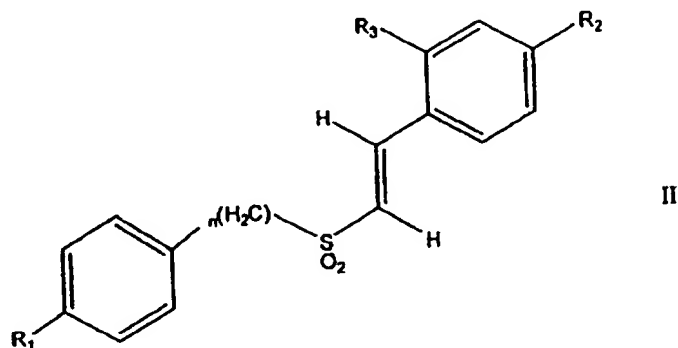


6. A compound according to claim 1 wherein the compound is E-4-chlorostyryl 4-bromobenzyl sulfone.

7. A compound according to claim 1 wherein the compound is E-4-bromostyryl 4-chlorobenzyl sulfone.

5 8. A compound according to claim 1 wherein the compound is E-4-bromostyryl 4-fluorobenzyl sulfone.

9. A pharmaceutical composition comprising a pharmaceutically acceptable carrier and a compound of the formula II



wherein

10 n is zero or one;  
 $R_1$  is selected from the group consisting of hydrogen, chlorine, fluorine and bromine;

$R_2$  is selected from the group consisting of hydrogen, chlorine, fluorine, bromine, methyl and methoxy; and

15  $R_3$  is selected from the group consisting of hydrogen, chlorine and fluorine;

provided,

$R_2$  may not be methyl or methoxy when  $R_1$  and  $R_3$  are both hydrogen and n is zero or one; and

20  $R_1$ ,  $R_2$  and  $R_3$  may not all be hydrogen when n is one.

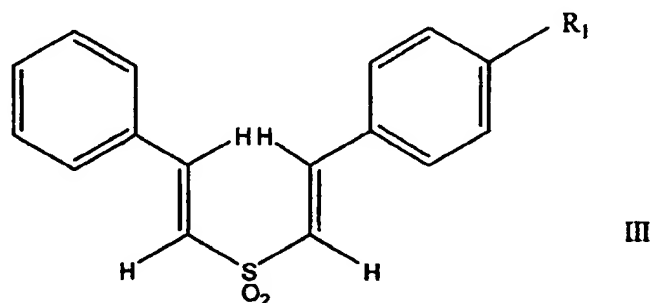
AMENDED SHEET (ARTICLE 19)

20. A composition according to claim 12 wherein the compound is E-4-bromostyryl 4-fluorobenzyl sulfone.

21. A composition according to claim 12 wherein the compound is E-4-chlorostyryl 4-bromobenzyl sulfone.

5 22. A composition according to claim 12 wherein the compound is E-4-bromostyryl 4-chlorobenzyl sulfone.

23. A pharmaceutical composition comprising a pharmaceutically acceptable carrier and a compound of the formula III

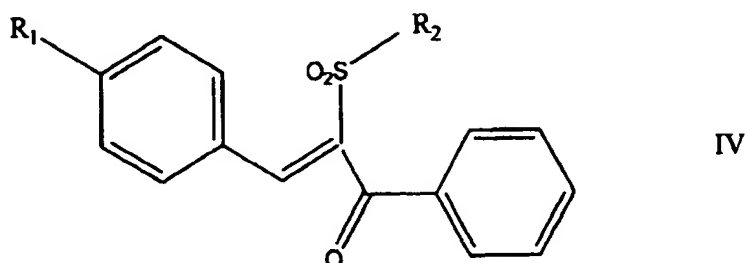


wherein

10

R<sub>1</sub> is selected from the group consisting of hydrogen, chlorine, fluorine and bromine.

24. A pharmaceutical composition comprising a pharmaceutically acceptable carrier and the compound of formula IV



wherein

5         $R_1$  is selected from the group consisting of fluorine and bromine, and  $R_2$  is selected from the group consisting of 2-chlorophenyl, 4-chlorophenyl, 4-fluorophenyl and 2-nitrophenyl.

25. A method of treating an individual for cancer comprising administering to said individual an effective amount of a pharmaceutical composition according to claim 9.

10        26. A method of inhibiting growth of tumor cells in an individual afflicted with cancer comprising administering to said individual a pharmaceutical composition according to claim 9.

15        27. A method of inducing apoptosis of tumor cells in an individual afflicted with cancer comprising administering to said individual a pharmaceutical composition according to claim 9.

28. A method of treating an individual for cancer comprising administering to said individual an effective amount of a pharmaceutical composition according to claim 23 or 24.

AMENDED SHEET (ARTICLE 19)

29. A method of inhibiting growth of tumor cells in an individual afflicted with cancer comprising administering to said individual a pharmaceutical composition according to claim 23 or 24.

5 30. A method of inducing apoptosis of tumor cells in an individual afflicted with cancer comprising administering to said individual a pharmaceutical composition according to claim 23 or 24.

31. A method according to claim 25 for treating breast or prostate cancer.

32. A method according to claim 26 for inhibiting growth of breast or prostate tumor cells in an individual afflicted with breast or prostate cancer.

10 33. A method according to claim 27 for inducing apoptosis of breast or prostate tumor cells in an individual afflicted with breast or prostate cancer.

34. A method according to claim 28 for treating breast or prostate cancer.

35. A method according to claim 29 for inhibiting growth of breast or prostate tumor cells in an individual afflicted with breast or prostate cancer.

15 36. A method according to claim 30 for inducing apoptosis of breast or prostate tumor cells in an individual afflicted with breast or prostate cancer.

AMENDED SHEET (ARTICLE 19)

1 / 7

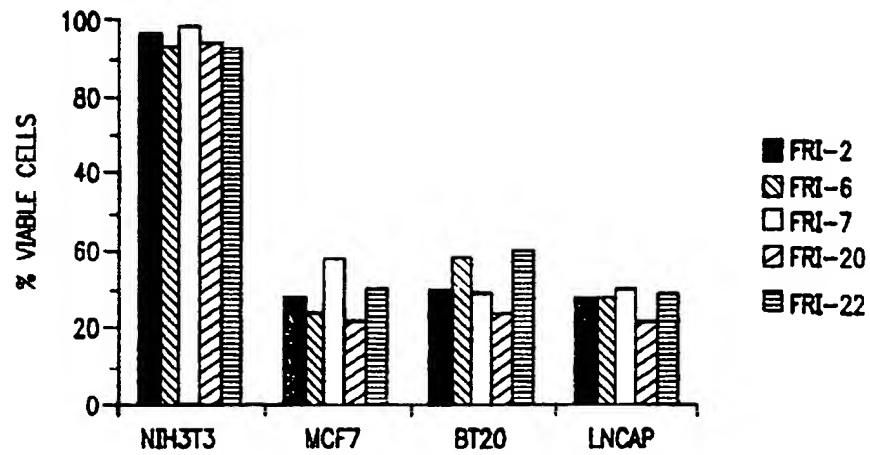


FIG. 1A

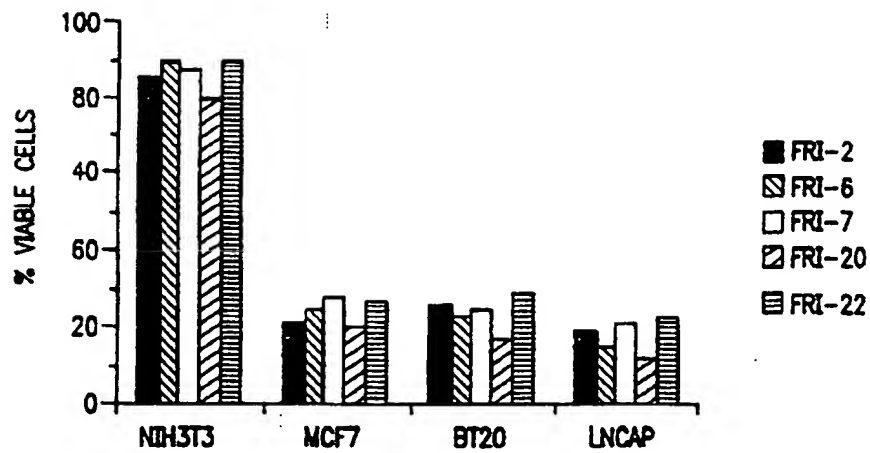


FIG. 1B

2/7

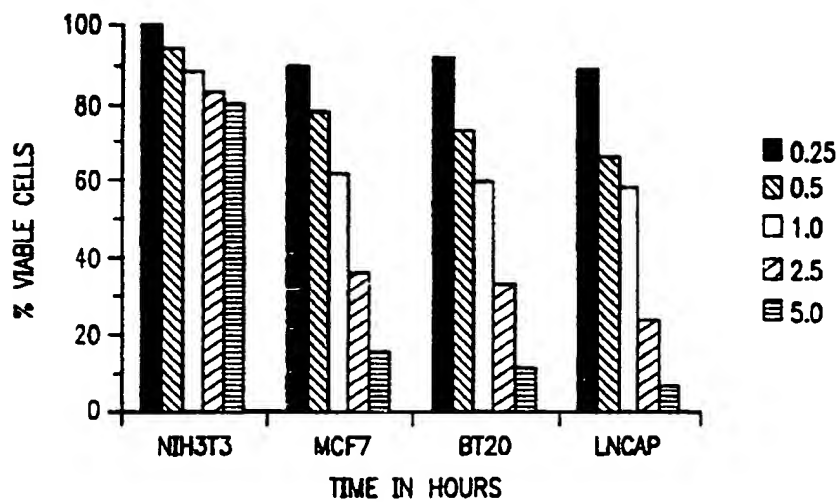


FIG. 2A

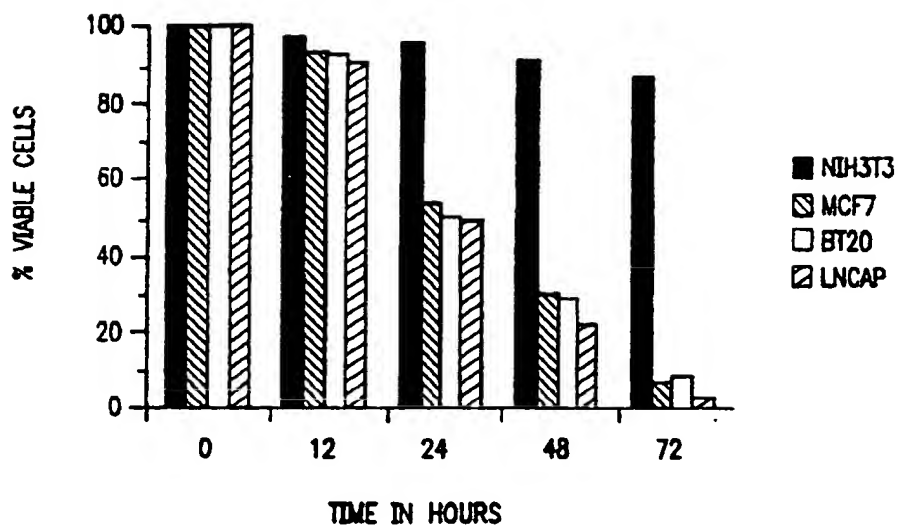


FIG. 2B

3/7

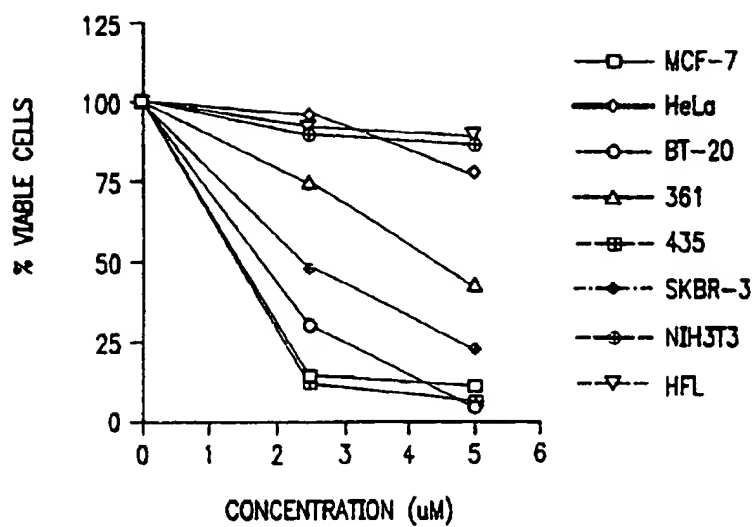


FIG. 3A

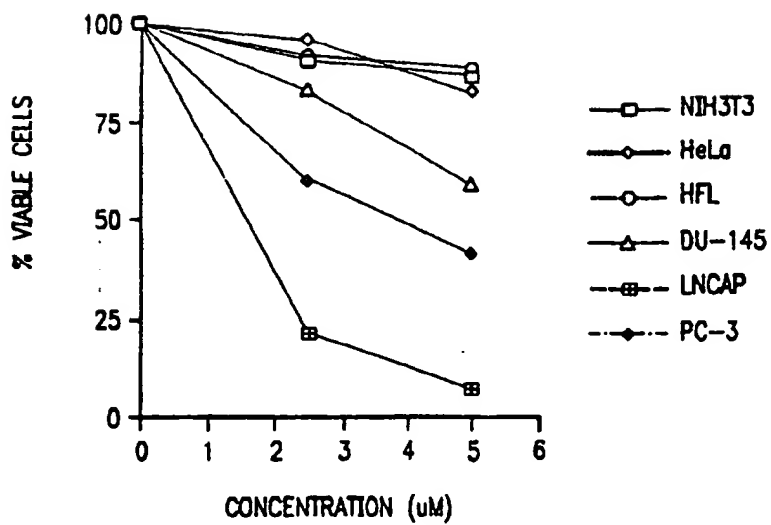


FIG. 3B

SUBSTITUTE SHEET (RULE 26)

4/7

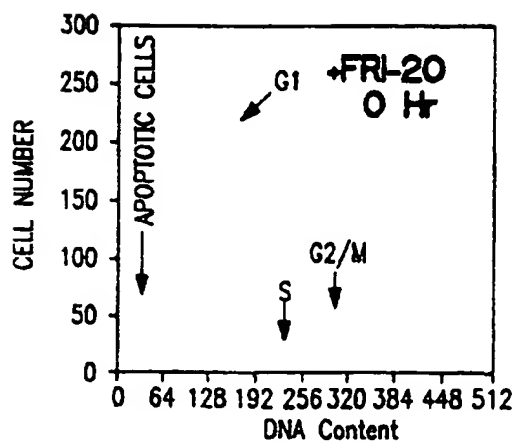


FIG. 4A

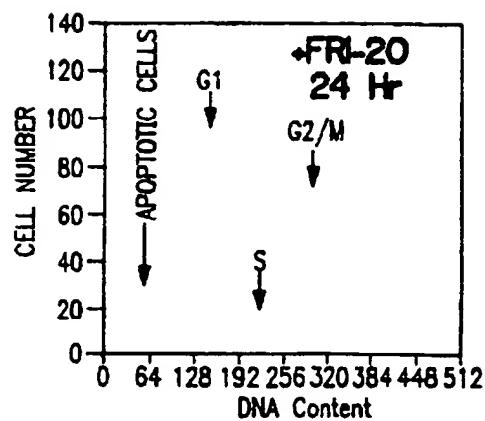


FIG. 4B

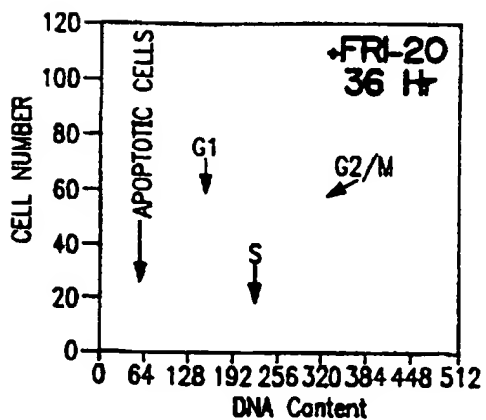


FIG. 4C

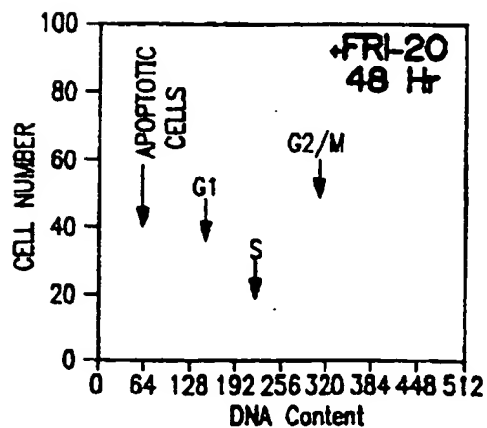


FIG. 4D



5/7

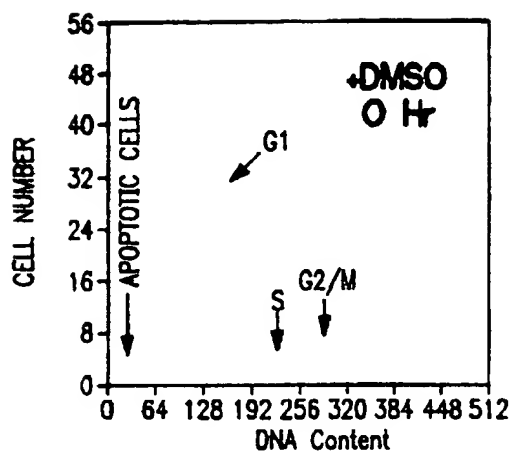


FIG. 4E

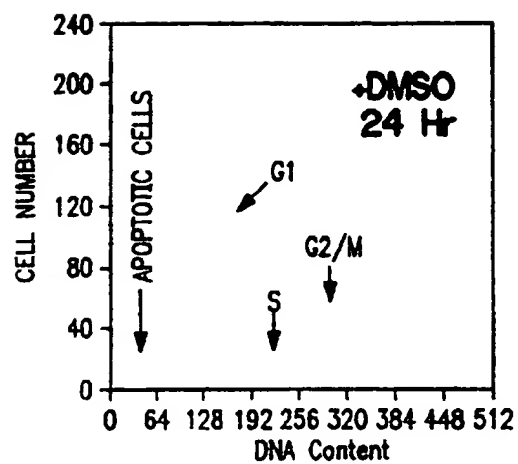


FIG. 4F

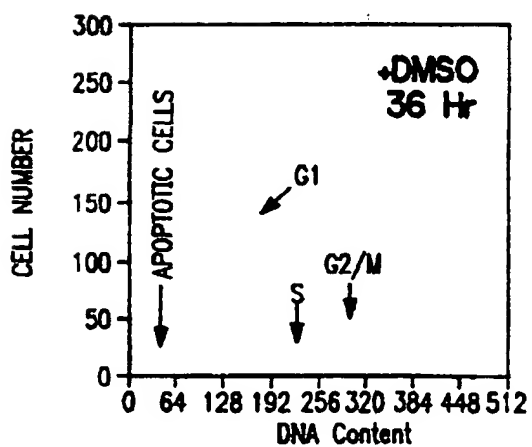


FIG. 4G

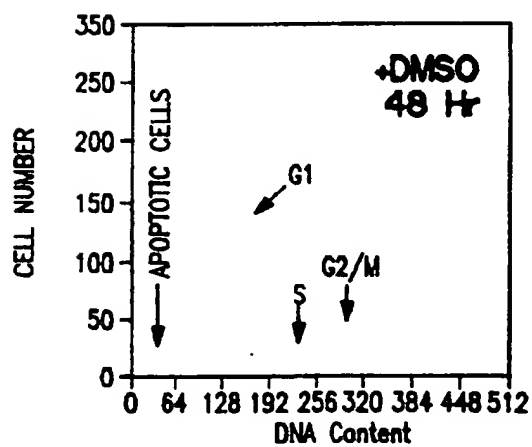


FIG. 4H

6/7

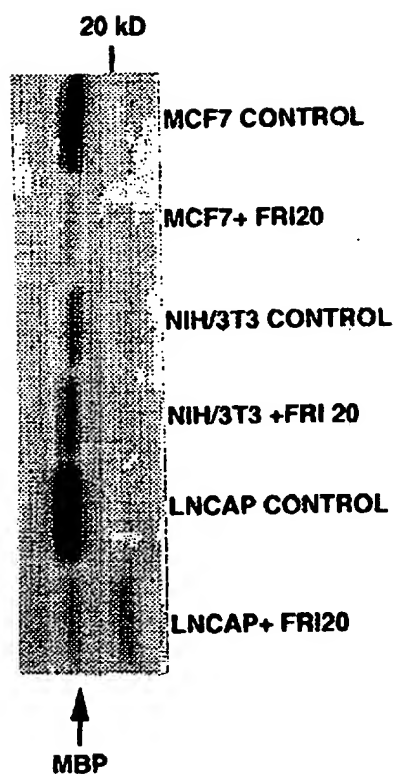


FIG. 5

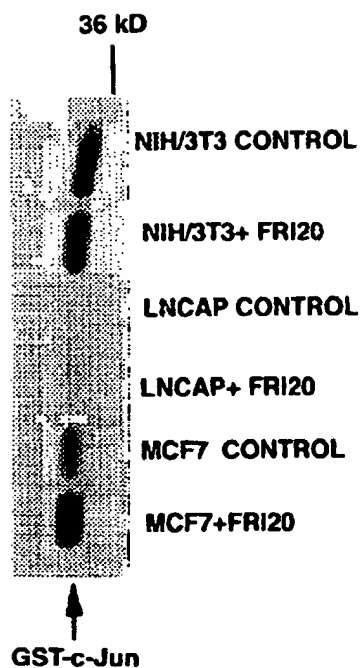
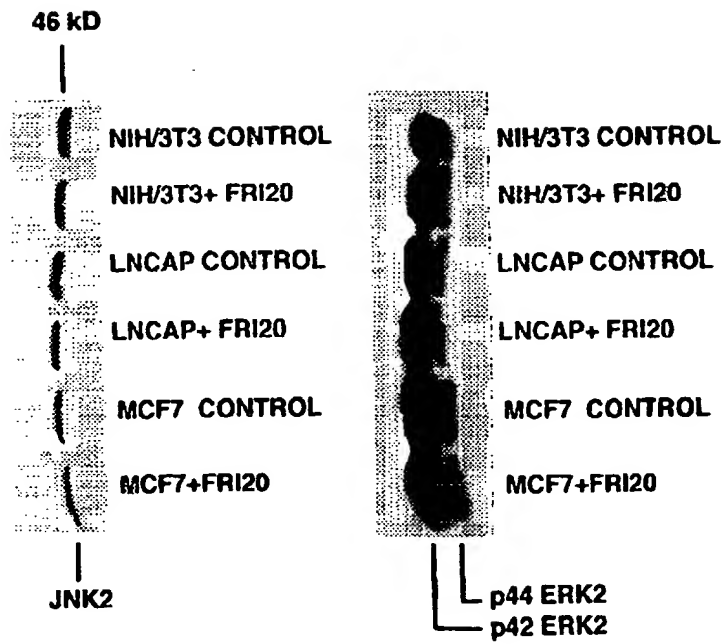


FIG. 7

7/7



**FIG. 6**

## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US98/20580

## A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : C07C 317/02, 317/14

US CL : 568/28, 31, 34, 35

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 568/28, 31, 34, 35

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched  
HAWLEY'S CHEMICAL DICTIONARY, 12th EditionElectronic data base consulted during the international search (name of data base and, where practicable, search terms used)  
Please See Extra Sheet.

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	Database Caplus on STN, Chem Abstracts, (Columbus OH, USA) No. 124:175763, REDDY, D.B. et al., 'Synthesis of 1,1-disubstitued 2,6-diaryl-4-thian-4,4-dioxides, part-II' abstract Indian J. Heterocycl. Chem., May 1995, Vol. 5, No. 1, pages 11-14.	1-24
A	Database Caplus on STN, Chem Abstracts, (Columbus OH, USA) No. 124:146025, REDDY, D.B. et al., 'E,Z and E,E- bis(styryl) sulfones as precursors of thiane oxides' abstract Indian J. Heterocycl. Chem. April 1995, Vol. 4, No. 4, pages 259-264.	1-24
A	US 4,937,388 A (BUSHELL et al) 26 June 1990 (26.06.90) column 1, lines 20-32 and abstract.	1-22

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	* Inter document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
* A* document defining the general state of the art which is not considered to be of particular relevance	* X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
* B* earlier document published on or after the international filing date	* Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
* L* document which may throw doubt on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	* A* document member of the same patent family
* O* document referring to an oral disclosure, use, exhibition or other means	
* P* document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

27 NOVEMBER 1998

Date of mailing of the international search report

29 DEC 1998

Name and mailing address of the ISA/US  
Commissioner of Patents and Trademarks  
Box PCT  
Washington, D.C. 20231

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Authorized Officer

JEAN F VOLLANO

Telephone No. (703) 308-1235

# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US98/20580

## C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A,P	Database Caplus on STN, Chem Abstracts (Columbus OH, USA) No. 126:166162, Thompson, H. et al. 'Sulfone metabolite of sulindac inhibits mammary carcinogenesis' abstract Cancer Res., February 1997, Vol. 57, No. 2, pages 267-271.	25-30

Form PCT/ISA/210 (continuation of second sheet)(July 1992)\*

# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US98/20580

## B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

APS, CAS ONLINE, BEILSTEIN

search terms: breast cancer, prostate cancer, sulfone, malignant tumor, structure drawings